STOCK SOLUTIONS

Bouin's Solution - see recipe in "Fixatives". Add glacial acetic acid (to 5%) just before use.

Alcoholic Hematoxylin Stock (Solution A)

<u>Component</u>	<u>Cat. No.</u>	\underline{Amount}
 Hematoxylin 	Sigma H9627	5 g
• 95% Ethanol		500 mL

Dissolve over gentle heat – do not boil. Label with date and initials. Store at room temperature up to 1 year.

10% Ferric Chloride Stock (Solution B)

<u>Compo</u>	<u>nent</u>	<u>Cat. No.</u>	<u>Amount</u>
•	Ferric chloride (Iron III chloride)	Sigma F7134	5.8 g
•	Milli-Pure water		495 ml
•	Glacial Acetic Acid		5 ml

Dissolve ferric chloride in 50 ml water, then add remaining water to 990 ml. Add acetic acid last. Store at room temperature for up to 1 year.

Acid Fuchsin, 1% Aqueous Stock Solution

Dissolve 1 g acid fuchsin (Sigma #857408) in 100 mL Milli-Pure water. Label and store at room temperature.

Biebrich Scarlet, 1% Aqueous Stock Solution

Dissolve 10 g biebrich scarlet (aka Ponceau BS, Sigma #B6008) in 1000 mL Milli-Pure water. Label and store at room temperature.

Aniline Blue Stock Solution

<u>Component</u>	To make 50 mL	To make 75 mL	To make 250 mL
• Aniline Blue (CAS #66687-07-8)	$1.25 \mathrm{~g}$	$1.875~\mathrm{g}$	$6.25~\mathrm{g}$
 Glacial acetic acid 	1.0 mL	$1.5~\mathrm{mL}$	5.0 mL
• DI water	50 mL	$75~\mathrm{mL}$	$250~\mathrm{mL}$

Store at room temperature for up to 1 year.

Phosphomolybdic-Phosphotungstic Acid Solution

<u>Component</u>	To make 50 mL	To make 75 mL	<u>To make 250 mL</u>
• Phosphomolybdic acid (Sigma #221856) 0.625 g	0.938 g	$3.125 \mathrm{g}$
• Phosphotungstic acid (Sigma #P4006)	$0.625 \mathrm{~g}$	0.938 g	$3.125 \mathrm{g}$
• DI Water	50 mL	$75~\mathrm{mL}$	$250~\mathrm{mL}$

Store at room temperature for up to 1 year.

WORKING SOLUTIONS

1% Acetic Acid Solution

<u>Component</u>	To make 50 mL	To make 75 mL	T <u>o make 250 mL</u>
Glacial acetic acid	0.5 mL	0.75 mL	$2.5~\mathrm{mL}$
Distilled Water	$49.5~\mathrm{mL}$	$74.25~\mathrm{mL}$	$247.5~\mathrm{mL}$

Make fresh just before use.

Weigert's Hematoxylin – Working Solution Lillehei Heart Institute – Histology & Microscopy Research Facility

<u>Compo</u>	\underline{nent}	To make 50 mL	To make 75 mL	<u>To make 250 mL</u>
•	Solution A – Alcoholic Hematoxylin Stock	k 25 mL	37.5 mL	$125~\mathrm{mL}$
•	Solution B – 10% Ferric Chloride Stock	25 mL	37.5 mL	$125~\mathrm{mL}$
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Prepare just before use. Discard in "Hematoxylin Waste" container.

Biebrich Scarlet-Acid Fuchsin Solution - Working Solution

<u>Component</u>	To make 50 mL	To make 75 mL	To make 250 mL
 Biebrich scarlet, 1% stock 	44.5 mL	66.75 mL	222.5 ml
 Acid fuchsin, 1% stock 	$5.0~\mathrm{mL}$	$7.5~\mathrm{mL}$	25 ml
Glacial Acetic Acid	0.5 mL	0.75 mL	2.5ml

Prepare just before use. Can be used for a maximum of two runs if within 24 hours.

Aniline Blue Working Solution

<u>Component</u>

•	Aniline Blue Stock Solution	50 ml	75 ml	250 ml
•	Glacial acetic acid	1.0 ml	1.5 ml	5.0 ml

Add glacial acetic acid just before using.

PROCEDURE:

Always run a control slide with every stain!

1. Deparaffinize and re-hydrate tissues to PBS*

2. Distilled water* 1 min.

3. Mordant in Bouin's – in fume hood, covered Overnight

4. Running tap water rinse – until yellow color clears 1 min.

5. Distilled water rinse 1 min.

6. Weigert's Hematoxylin working solution* 12 min.

a. Discard after use.

7. Running tap water rinse 10 min.

8. Distilled water rinse 1 min.

9. Biebrich scarlet-acid fuchsin solution 1 min.

a. Solution may be used twice only, then discarded.

10. Distilled water rinse 1 quick dip only

11. Phosphomolybdic-phosphotungstic acid solution 30 min.

a. Discard after use.

12. Aniline blue solution*

a. Solution may be used twice only, then discarded.

13. Distilled water rinse 1 quick dip only

14. 1% Acetic acid solution 4 min.

a. Discard after use.

15. 95% Ethanol 2 x 1 min.

16. 100% Ethanol 2 x 1 min.

17. Formula 83 3 x 1 min.

18. Mount with Permount

*MODIFICATIONS FOR CRYO-SECTIONS:

- 1. Fix sections in 10%formalin (not Zinc) x 30 minutes.
- 2. Replace steps 1, 2 with 3 x 3 min. rinses in distilled water.
- 3. Reduce time in hematoxylin and aniline blue to 7 minutes each.

RESULTS:Cytoplasm, keratin, muscle fibers, Erythrocytes – red

Nuclei - black

Collagen and mucus - blue

REFERENCES:

Carson, Freida L., Histology, A Self-Instructional Text. 1990. Pp. 142-144, and Plate 8-4, p. 267.

As cited by the American Ukranian Medical Project at:

http://aump.org/resources/documents/histo/en/MassonsTrichrome.pdf