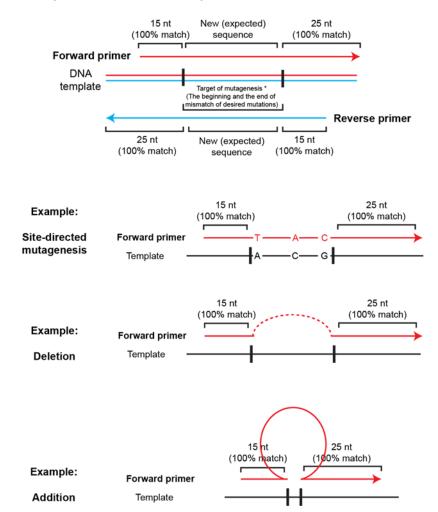
### Mutagenesis protocol (Jennifer Doudna lab)

- 1. The protocol is able to make site-directed mutagenesis for single or multiple nucleotides, introducing new sequences, and deleting a chunk of sequences.
- 2. Design the primers following the rules below.



- 3. Synthesize primers using standard desalting method (no extra purification needed)
- 4. Perform long-range PCR with PfuTurbo (Agilent # 600250), PfuUltra (Agilent # 600380) or PfuUltra II (Agilent # 600670).

#### 5. Condition for PfuTurbo:

## Optimization Parameters and Suggested Reaction Conditions (50 $\mu\text{l}$ reaction volume)

Parameter	Targets: <10 kb (vector DNA) or <6 kb (genomic DNA)	Targets: >10 kb (vector DNA) or >6 kb (genomic DNA)
Extension time	1 min per kb	2 min per kb
PfuTurbo DNA polymerase	2.5 U	5.0 U
Input template	50–100 ng genomic DNA°	200–250 ng genomic DNA°
Primers (each)	100-200 ng (0.2-0.5 μM)	200 ng (0.5 μM)
dNTP concentration	100–250 μM each dNTP (0.4–1.0 mM total)	500 μM each dNTP (2 mM total)
Final reaction buffer concentration	1.0×	1.5× (genomic DNA targets) 1.0× (vector DNA targets)
Denaturing temperature	95°C	92°C
Extension temperature	72°C	68°C

# Reaction Mixture for a Typical Single-Copy Chromosomal Locus PCR Amplification

Component	Amount per reaction
Distilled water (dH <sub>2</sub> O)	40.6 μΙ
10× cloned Pfu reaction buffer <sup>a</sup>	5.0 μl
dNTPs (25 mM each dNTP)	0.4 μΙ
DNA template (100 ng/µl)	1.0 µl <sup>b</sup>
Primer #1 (100 ng/μl)	1.0 μl <sup>c</sup>
Primer #2 (100 ng/μl)	1.0 μl <sup>c</sup>
PfuTurbo DNA polymerase (2.5 U/μl)	1.0 μl (2.5 U) <sup>d</sup>
Total reaction volume	50 μΙ

## A. Targets <10 kb (vector DNA) or <6 kb (genomic DNA)

Segment	Number of cycles	Temperature	Duration
1	1	95°℃	2 minutes
2	30	95°C	30 seconds
		Primer $T_m - 5^{\circ}C^{\circ}$	30 seconds
		72°C	1 minute for targets ≤1 kb
			1 minute per kb for targets >1 kb
3	1	72°C	10 minutes

## B. Targets >10 kb (vector DNA) or >6 kb (genomic DNA)

Segment	Number of cycles	Temperature	Duration
1	1	92°C	2 minutes
2	10	92°C	10 seconds
		Primer $T_m - 5^{\circ}\text{C}^d$	30 seconds
		68°C	2 minutes per kb
3	20	92°C	10 seconds
		Primer $T_m - 5^{\circ}\text{C}^d$	30 seconds
		68°C	2 minutes per kb plus 10 seconds/cycle